

# Tracking Tart Cherry Fruit Development

Anna Cohen<sup>1</sup>, Victoria Meakem<sup>2</sup>, Benjamin Gutierrez<sup>2</sup>

<sup>1</sup>College of Agriculture and Life Sciences, Cornell University, Ithaca NY, 14850

<sup>2</sup>Plant Genetic Resources Unit, USDA-ARS, Geneva, NY 14456



## Introduction

'Montmorency' is the primary cultivar of tart cherry (*Prunus cerasus*) grown in the United States, but there are many other varieties with commercial potential. The USDA Tart Cherry Collection (Geneva NY) contains 149 genetically distinct accessions, each with a unique profile of phenolic compounds and sugar-acid ratios that change over the growing season. Since cherries are non-climacteric fruit, they stop ripening once picked from the tree. Thus, growers must harvest at a time when fruit quality matches consumer preferences for sweetness, appearance, and health benefits derived from phenolic antioxidants. This project characterizes how 'Montmorency' and three lesser-known varieties ripen over three weeks by measuring sugar content, color, and phenolic compounds. Providing more information on lesser-known varieties could help the tart cherry industry diversify into novel cultivars.

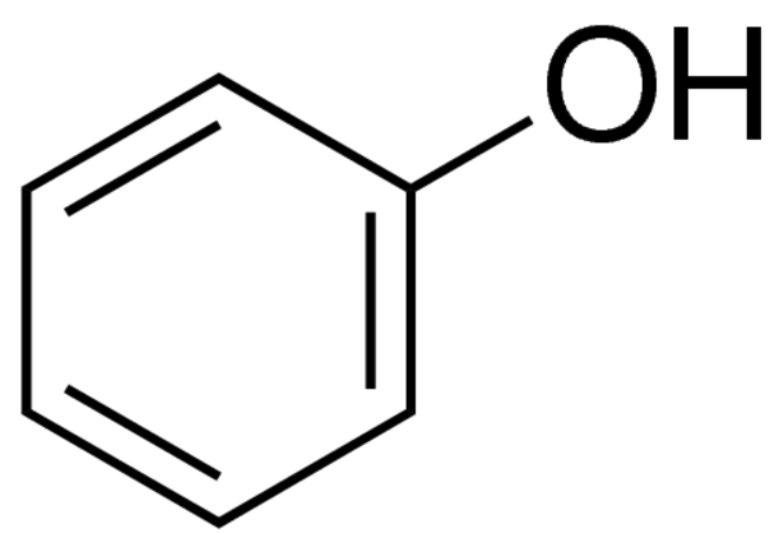


Figure 1: Phenol ring



Figure 2: USDA Tart Cherry Collection. Photo Credit: Zachary Stansell

## Materials and Methods

Harvested tart cherries on six dates from June 26 to July 13, 2023 from four varieties:

- 'Montmorency'
- 'Balaton'
- 'Fructbare von Michurin'
- 'Itt 18 (12)'



Figure 3: Colorimeter used to measure L\* a\* b\* values



Figure 4: Refractometer used to measure Brix (sugar percentage) on juice samples

Prepared frozen cherries samples to perform High Performance Liquid Chromatography (HPLC):

- Ground cherry flesh into powder
- Extracted 0.5 mg cherry with 1.5 mL of solvent (70 % MeOH, 2% CH<sub>2</sub>O<sub>2</sub> in water)
- Mixed, centrifuged, filtered.
- Ran samples with the lab's HPLC cherry phenolic protocol (Chang et al., 2010)

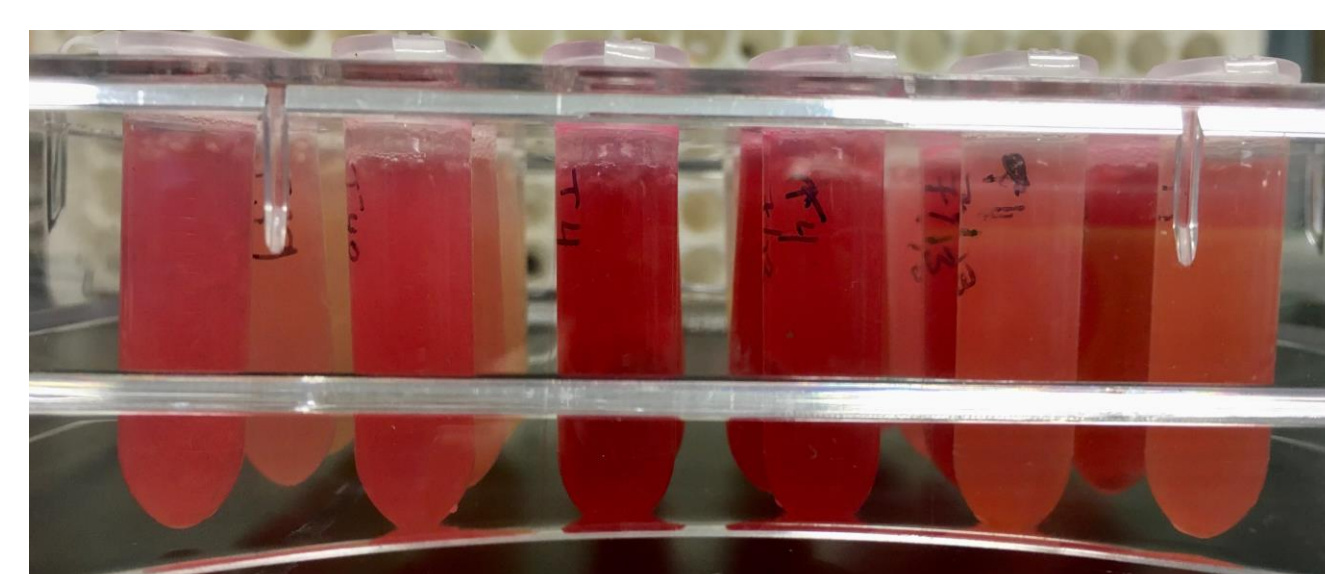


Figure 5: 2 mL tubes of extracted cherry flesh, prior to filtering

Major compounds were identified by matching retention time and wavelength with previous cherry phenolics lab results.

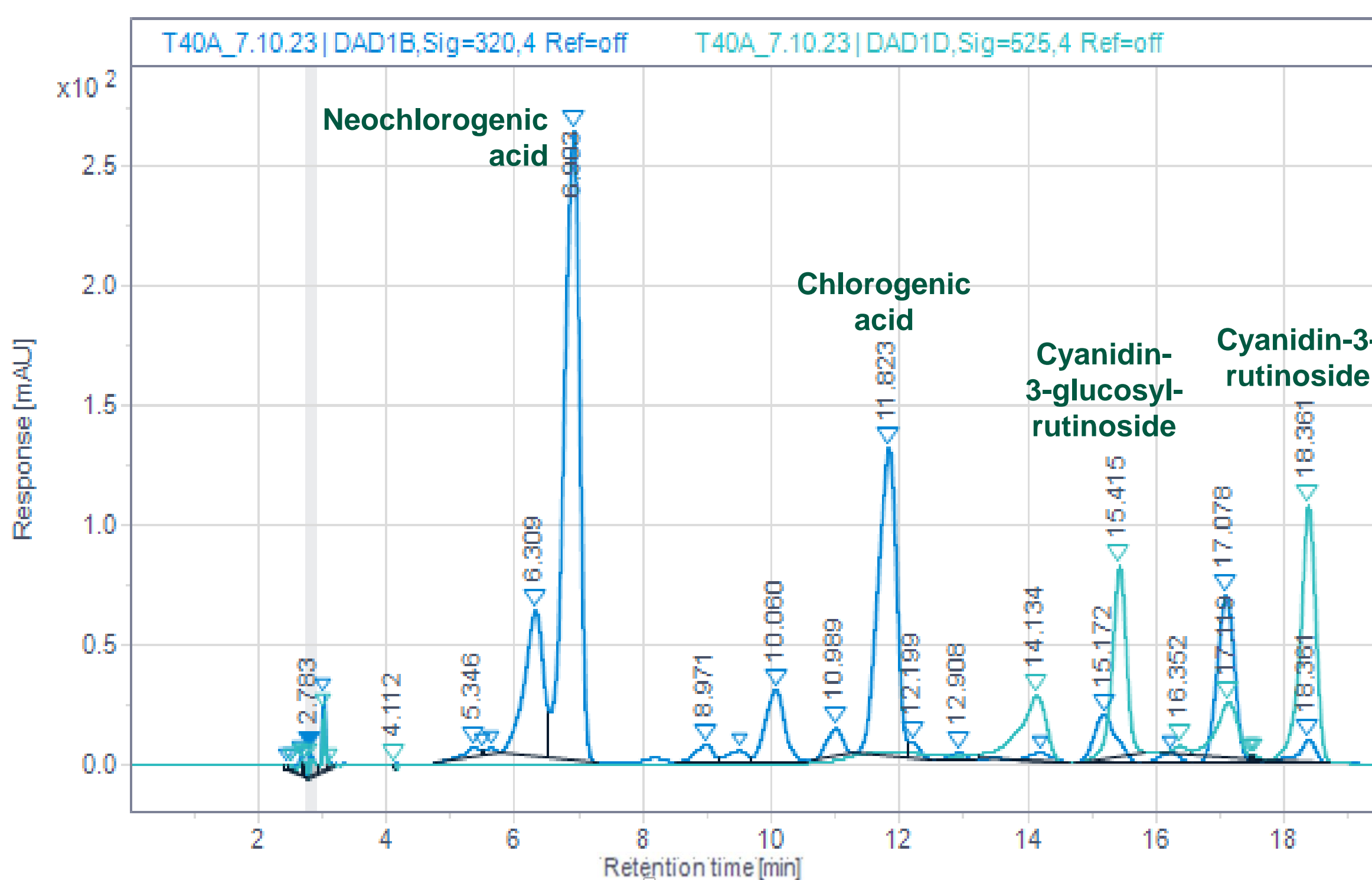


Figure 6: HPLC chromatogram for Itt 18 (12) harvested on June 10.

## References

- Kack, K. Maturation and picking time for sweet cherries (*Prunus avium*) and sour cherries (*Prunus cerasus* L.). *Eur Food Res Technol* 243, 539–546 (2017). <https://doi.org/10.1007/s00217-016-2753-6>
- Karaaslan, M., Yilmaz, F. M., Karaaslan, A. & Vardin, H. Synthesis and accumulation of anthocyanins in sour cherries during ripening in accordance with antioxidant capacity development and chalcone synthase expression. *Eur Food Res Technol* 242, 189–198 (2016).
- Sandra, P., Levaj, B., Verica, D., Škevin, D., Skendrovic, M. Color Parameters and Total Anthocyanins of Sour Cherries (*Prunus cerasus* L.) During Ripening. *Agric. conspec. sci.* 74, (2009).
- Zhang, Y., Li, P. & Cheng, L. Developmental changes of carbohydrates, organic acids, amino acids, and phenolic compounds in 'Honeycrisp' apple flesh. *Food Chemistry* 123, 1013–1018 (2010).

## Results

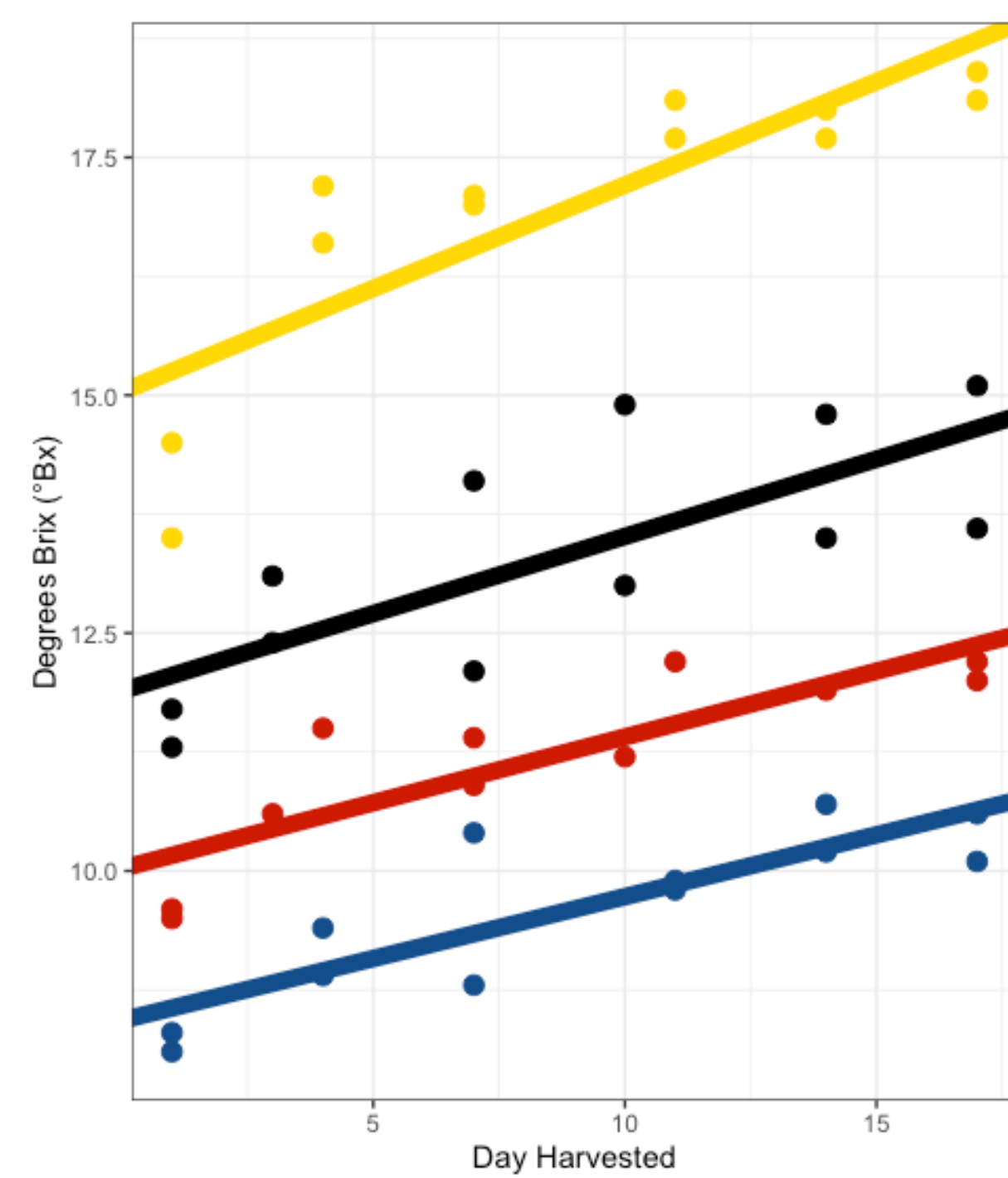


Figure 7: Brix (Sugar Percentage) Over Time

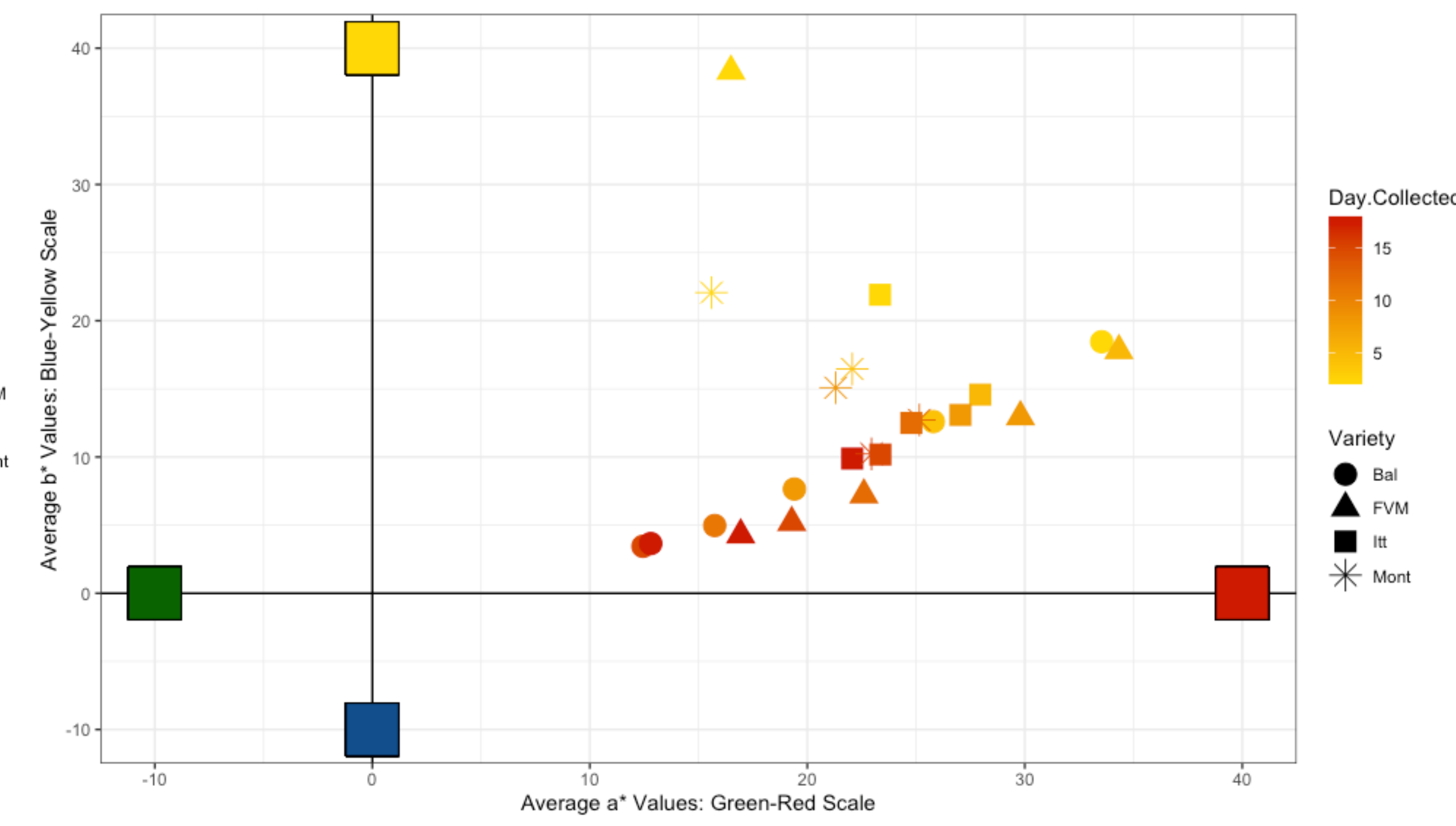


Figure 8: Colorimeter Hue Over Time

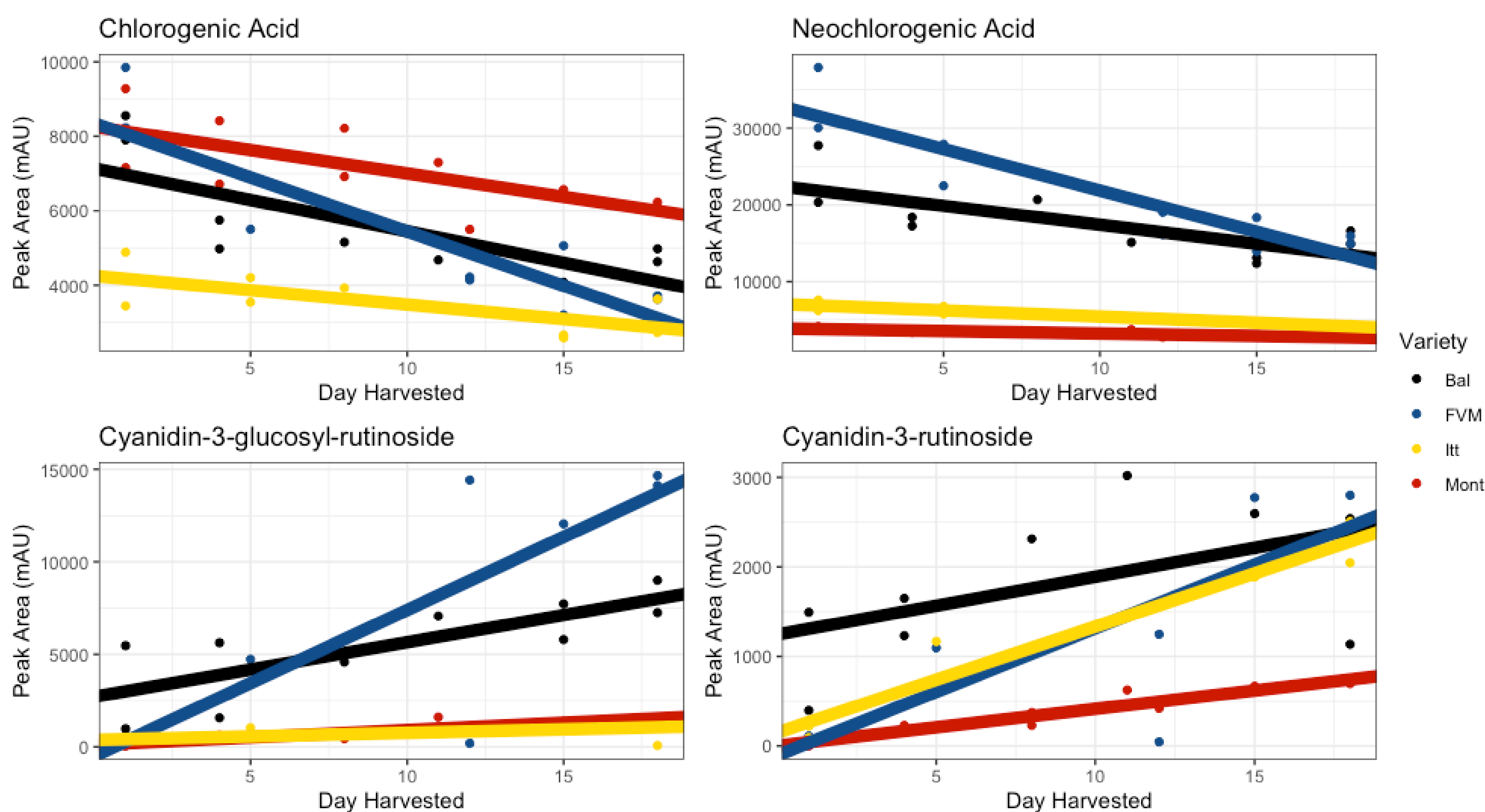


Figure 9: Phenolic Compounds Over Time in 'Balaton' (Bal), 'Fructbare von Michurin' (FVM), 'Itt 18 (12)' (Itt), 'Montmorency' (Mont)

Brix values for all varieties increased over time ( $p < 0.001$ ), and all varieties differed significantly from each other, (all  $p < 0.001$ ) (Fig. 7).

The cherries become more red and the color becomes less brilliant over time (Fig. 8). The lightness value ( $L^*$  value) decreases over time, demonstrating the cherries becomes darker (not pictured).

In all four phenolic compounds, the day harvested, variety, and their interaction were significant predictors of peak area (all  $p < 0.03$ ). This indicates that depending on cherry variety, the day harvested predicts peak area differently (see slopes in Fig. 9). Neochlorogenic acid and chlorogenic acid decreased over time for all varieties ( $p < 0.001$ ). Cyanidin-3-glucosyl-rutinoside and cyanidin-3-rutinoside increased over time for all varieties ( $p < 0.02$ ) (Fig. 9).

## Discussion and Future Work

Previous studies on tart cherry maturation investigated different varieties in different field locations. However, many trends remained consistent. Reports showed increases in Brix (Kack, 2017) and decreases in  $L^*$  values and brilliance (Sandra et al., 2009). Additionally, increases in cyanidin-3-glucosyl-rutinoside and cyanidin-3-rutinoside were published (Karaaslan et al., 2015).

Several HPLC peaks on the chromatograms remain unspecified (Fig. 6), so identifying these compounds would be the first step to a better understanding of the tart cherry profile.

Different tart cherry sweetness, color, and phenolic profiles may be matched to different markets (fresh, juicing, or canning). Developing full profiles for each variety may translate to better products. We recommend a broader study over a longer period of time, incorporating data from multiple field locations and seasons.

Developing a proxy indicator for phenolic compounds (e.g. Brix values that are easy to measure) would help growers identify when to harvest their fruit. With more data regarding ripening, it would become more feasible for growers to expand into tart cherry varieties beyond 'Montmorency.'

## Acknowledgments

We are grateful for the USDA NIFA Enhanced Workforce Development REEU grant 2021-68010-34652 that funded this project. We thank the Cornell Geneva Summer Scholars program for their additional support. We would like to thank Erin Galarneau, Dan Meyers, and Tony Barraco for technical assistance, as well as Marcella Venetozzi, Sam Page, Abe Porschet, Johnny Aponte, and Mica Patterson for assistance in sample processing.

